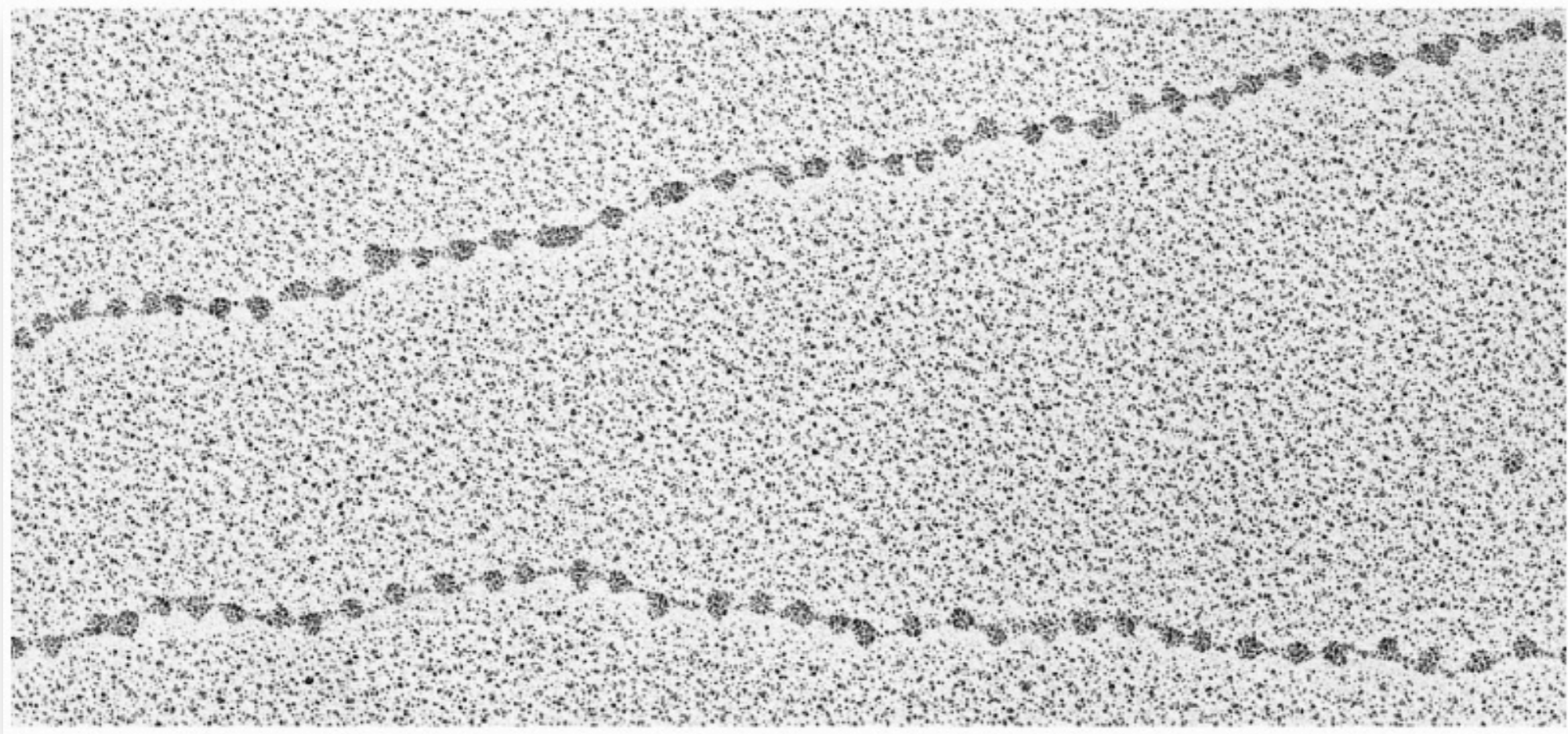


Chromatin organization and dynamics

1973: The first visualization of nucleosome chains by electron microscopy

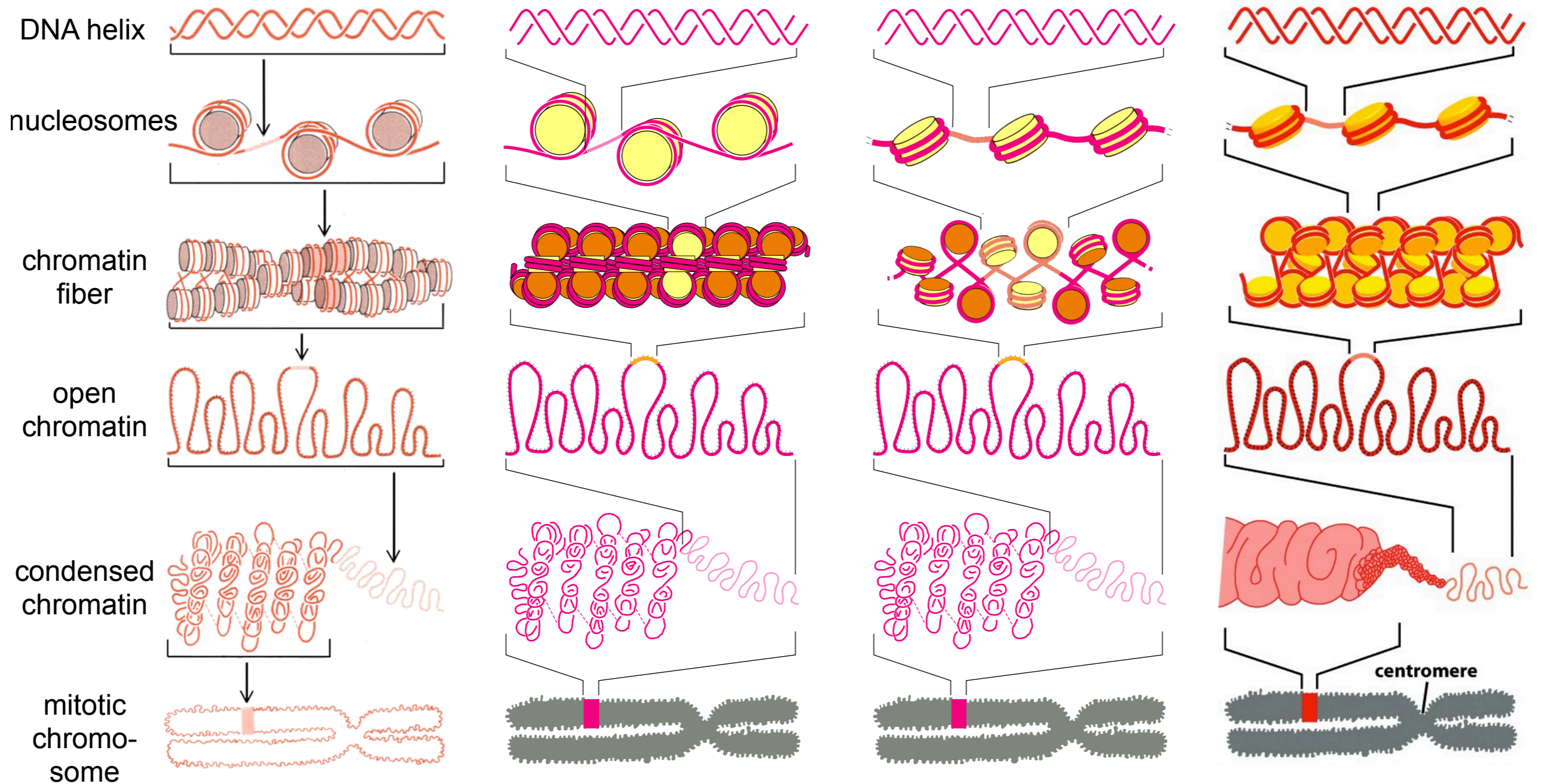


Olins, A. L. and Olins, D. E. (1974) *Science* 183, 330-332.

Reviewer's comment on the paper Chris Woodcock submitted in 1973:

“I have never read such a naive paper purporting to be of such fundamental significance. Definitely it should not be published anywhere!”

The last 25 years of chromatin structure research from the “Molecular Biology of the Cell” textbook



Alberts et al. 1983

Alberts et al. 1994

Alberts et al. 2002

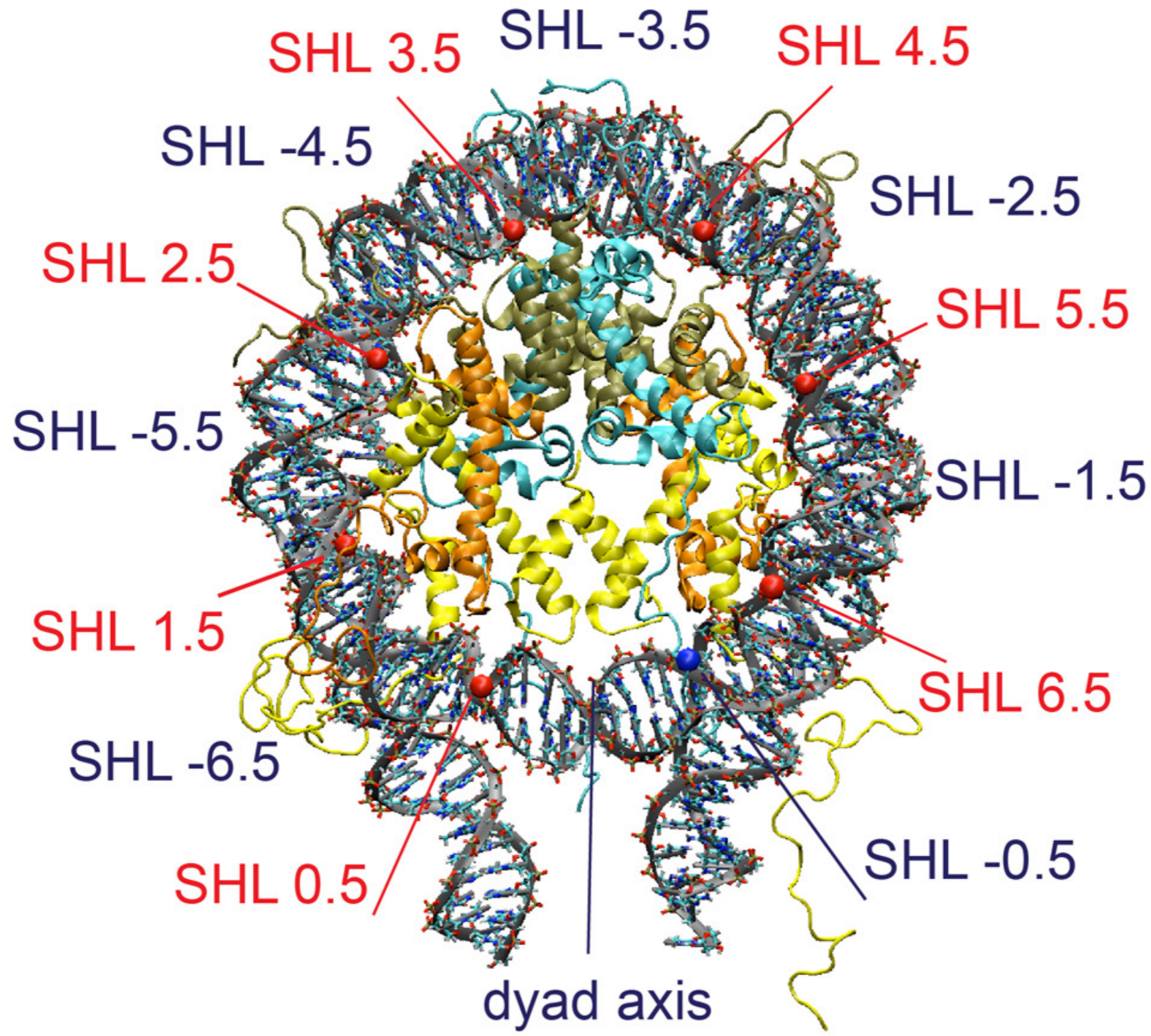
Alberts et al. 2007

Coarse-grained nucleosome and chromatin models

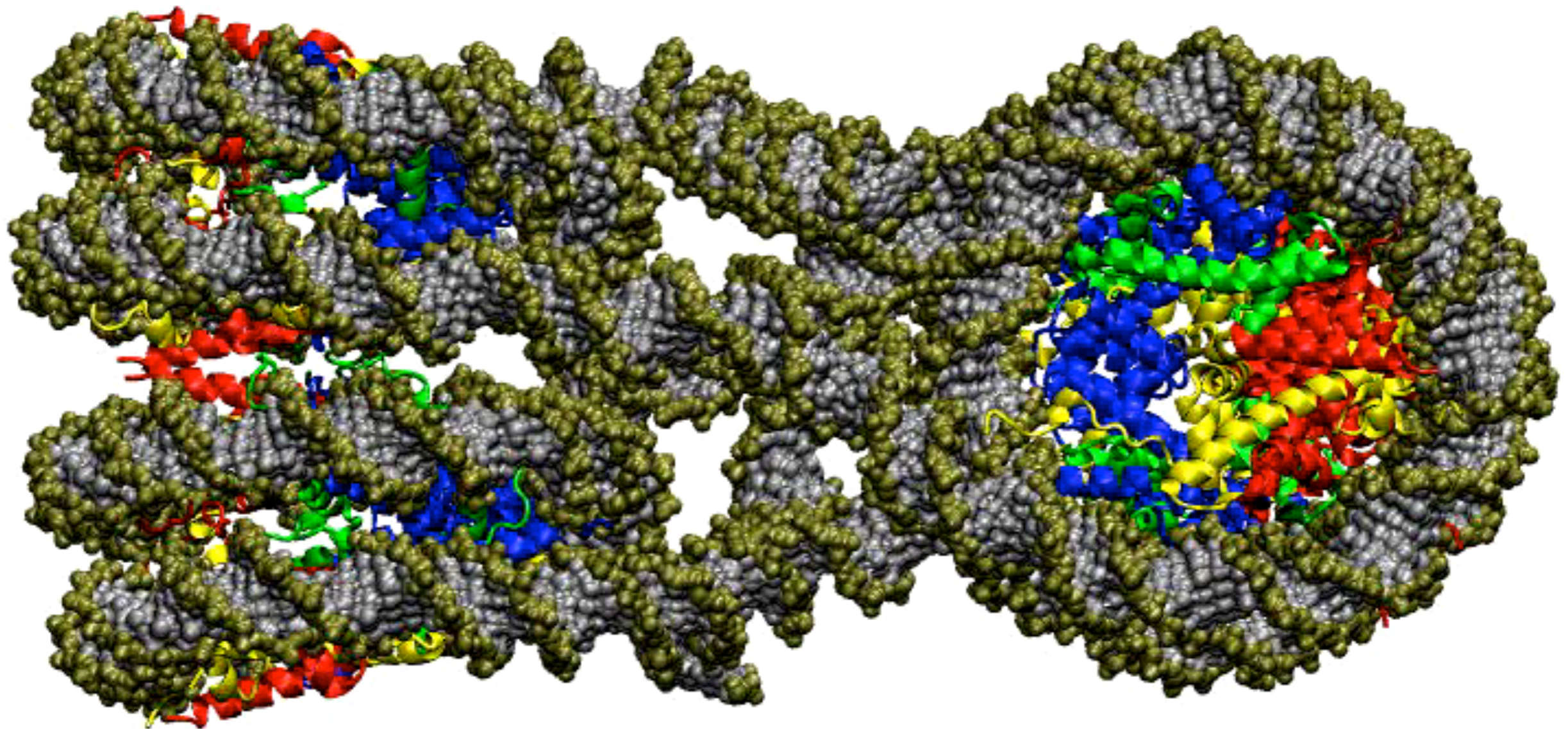
(Rippe et al., unpublished)



We know a lot about the nucleosome

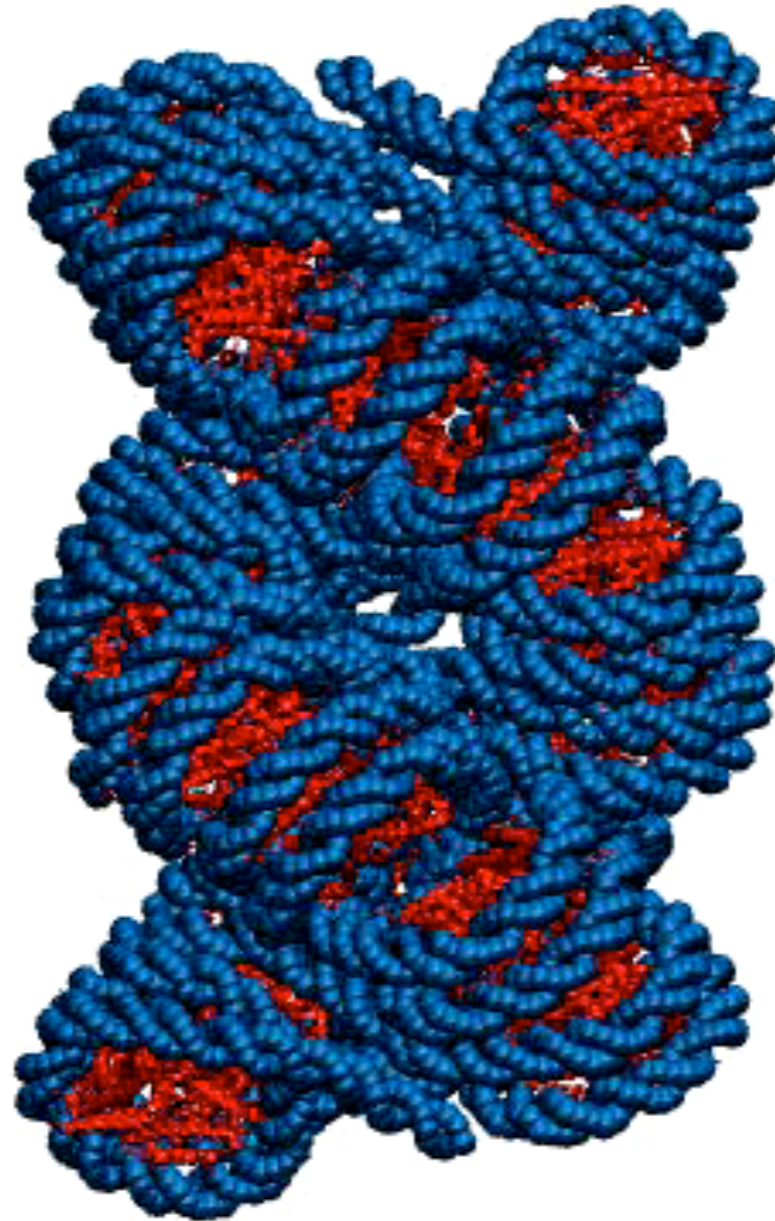


A tetranucleosome structure without H1
and a repeat length of 169 base pairs

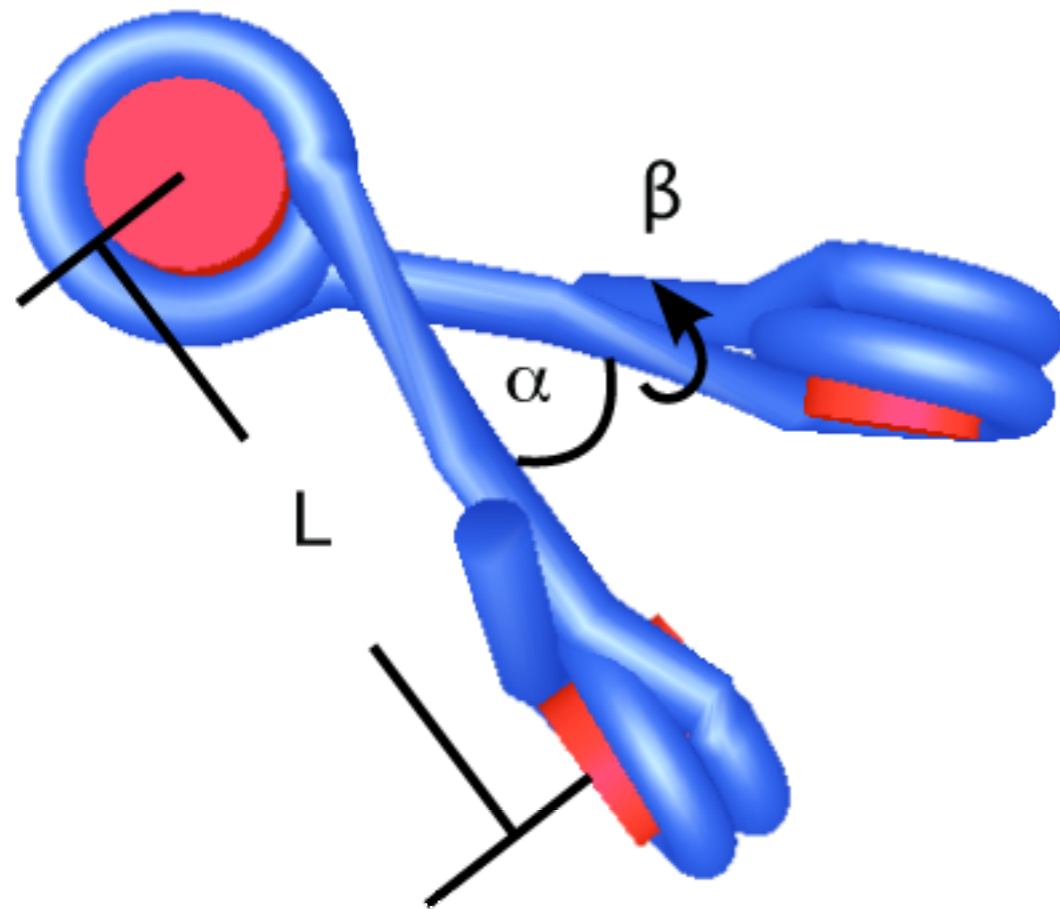


Schalch, T., Duda, S., Sargent, D.F., and Richmond, T.J., Nature, 2005

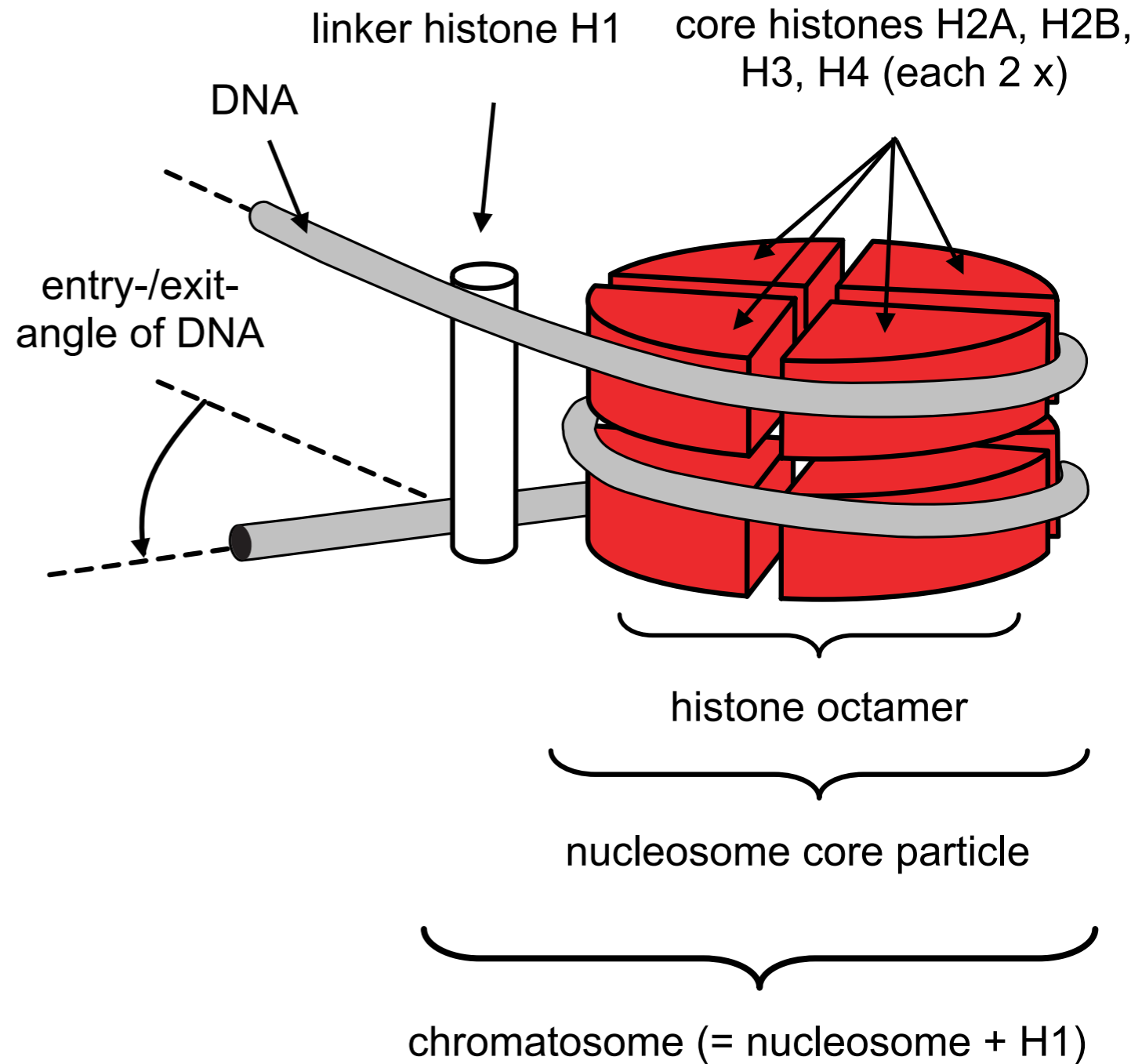
Can the tetra-nucleosome structure be extended into a stable chromatin fiber? And what about other repeat lengths?



The two-angle description of the nucleosome geometry



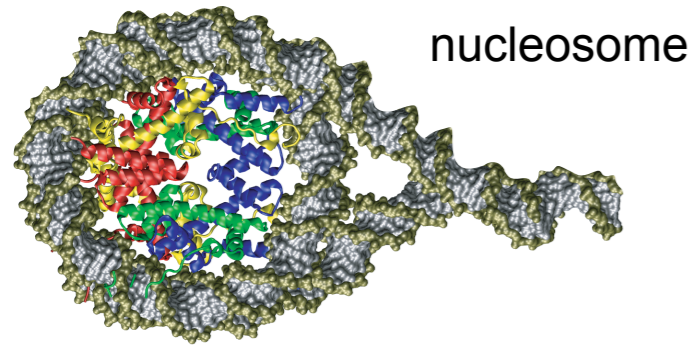
Woodcock, C.L. et al., PNAS, 1993



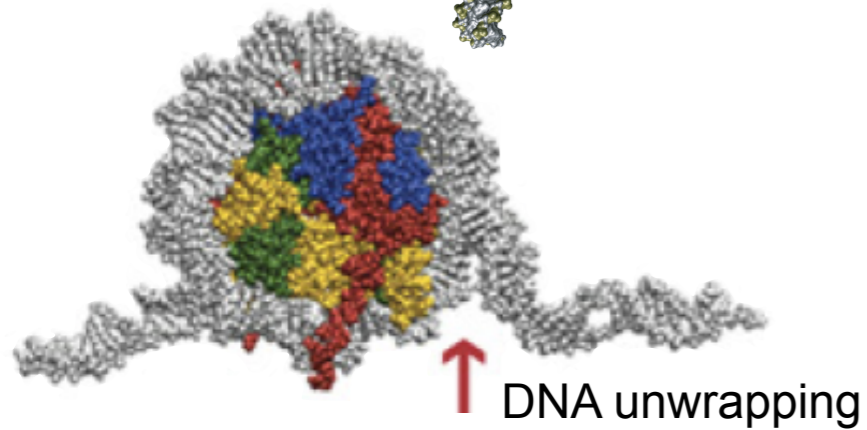
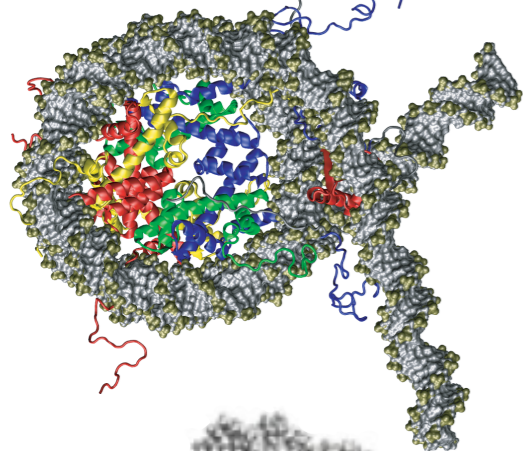
linker length L : length of DNA between nucleosomes
repeat length NRL: linker length + DNA around the nucleosome

The nucleosome chain conformation regulates DNA access

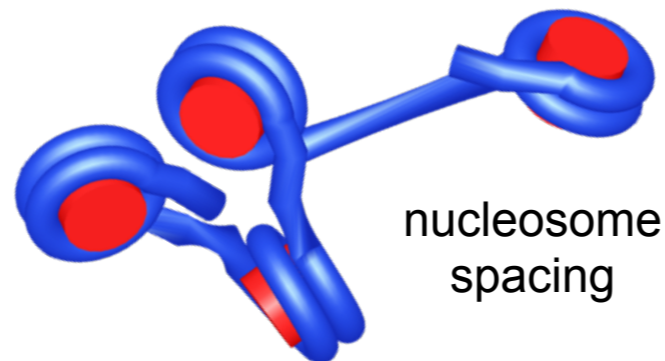
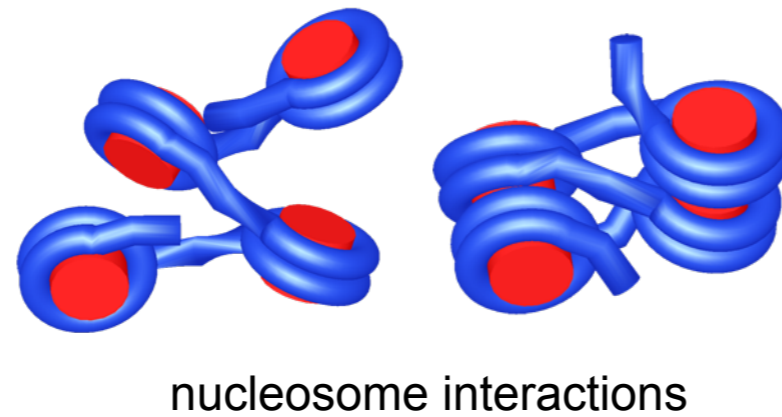
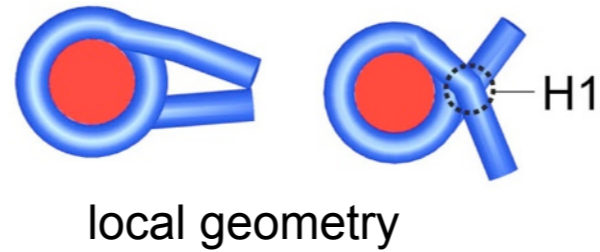
All atom models



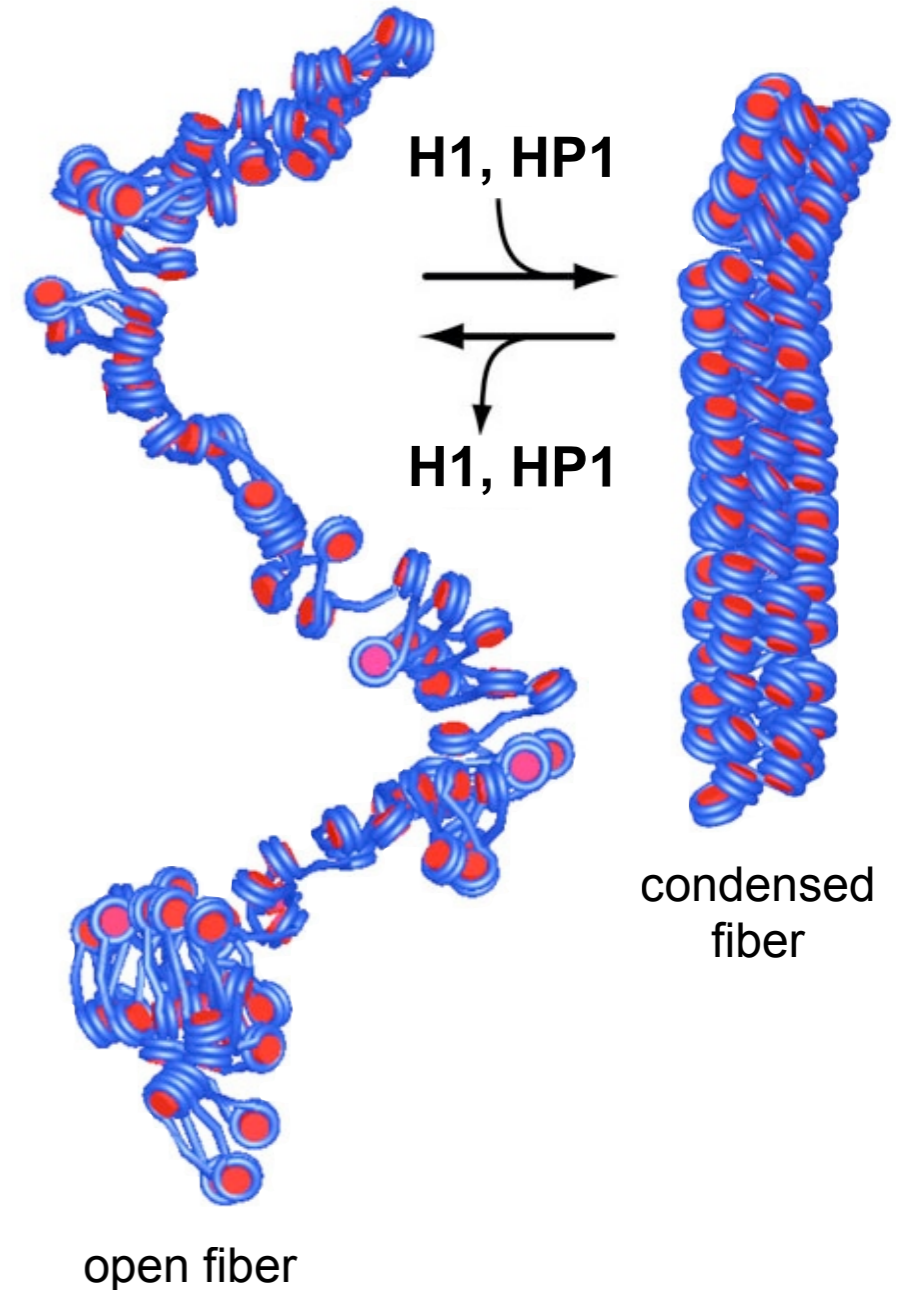
+ linker histone H1



Coarse-grained representation



Fiber conformation



Kepper, Foethke, Stehr, Wedemann & Rippe. *Biophys. J.* (2008)

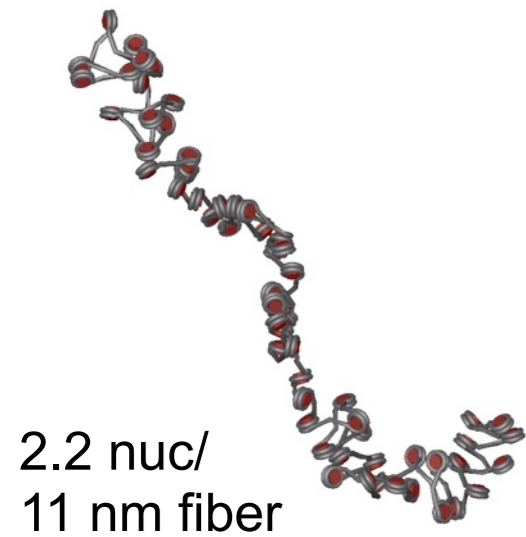
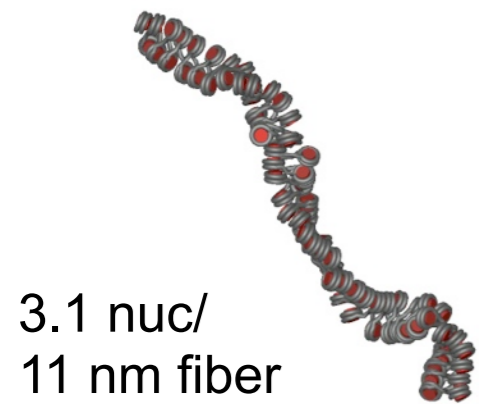
Teif, Ettig & Rippe, *Biophys. J.* (2010)

Ettig, Kepper, Stehr, Wedemann & Rippe, *Biophys. J.* (2011)

Kepper, N., Ettig, R., Stehr, R., Wedemann, G. & Rippe, K. *Biopolymers* (2011).

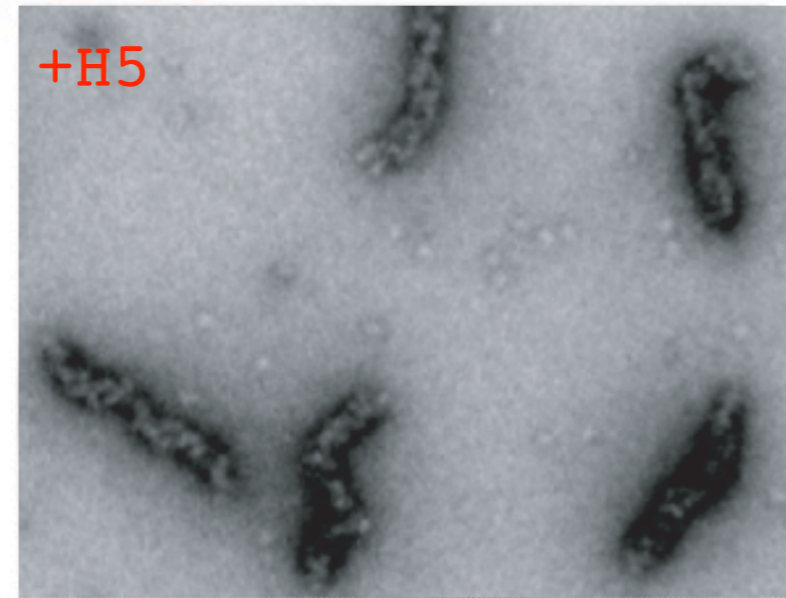
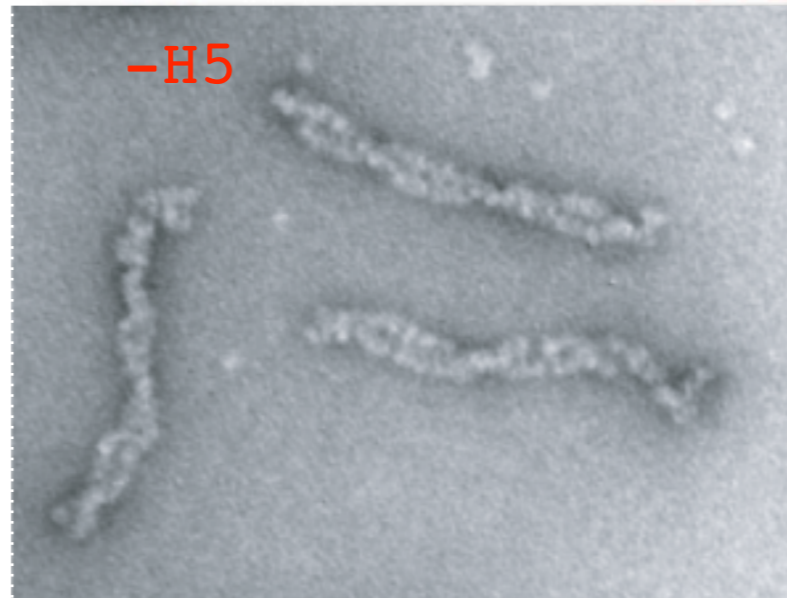
We understand some things about reconstituted nucleosome chains in vitro

MC simulations

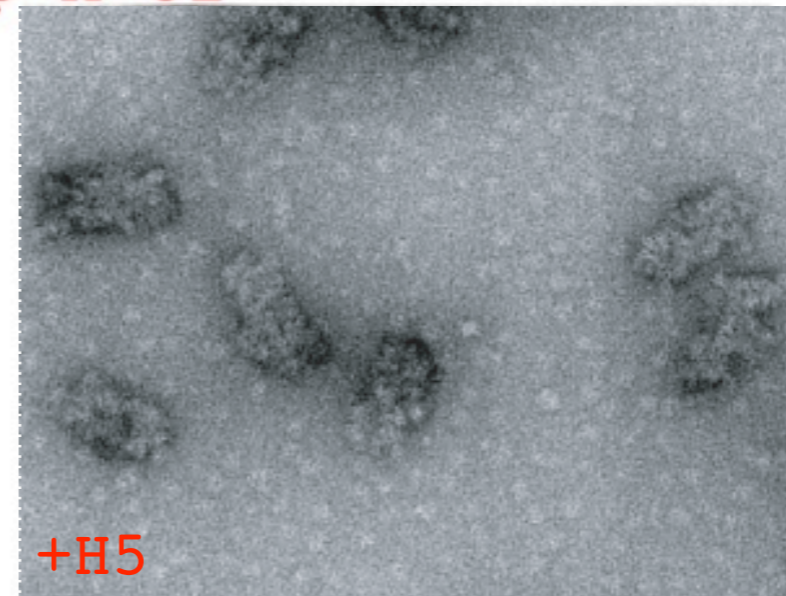
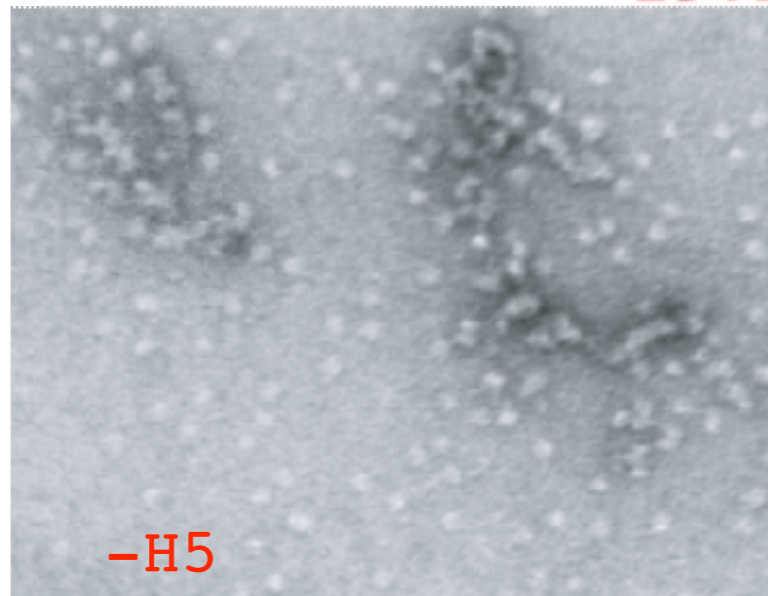


EM images of chromatin

167bp x 80



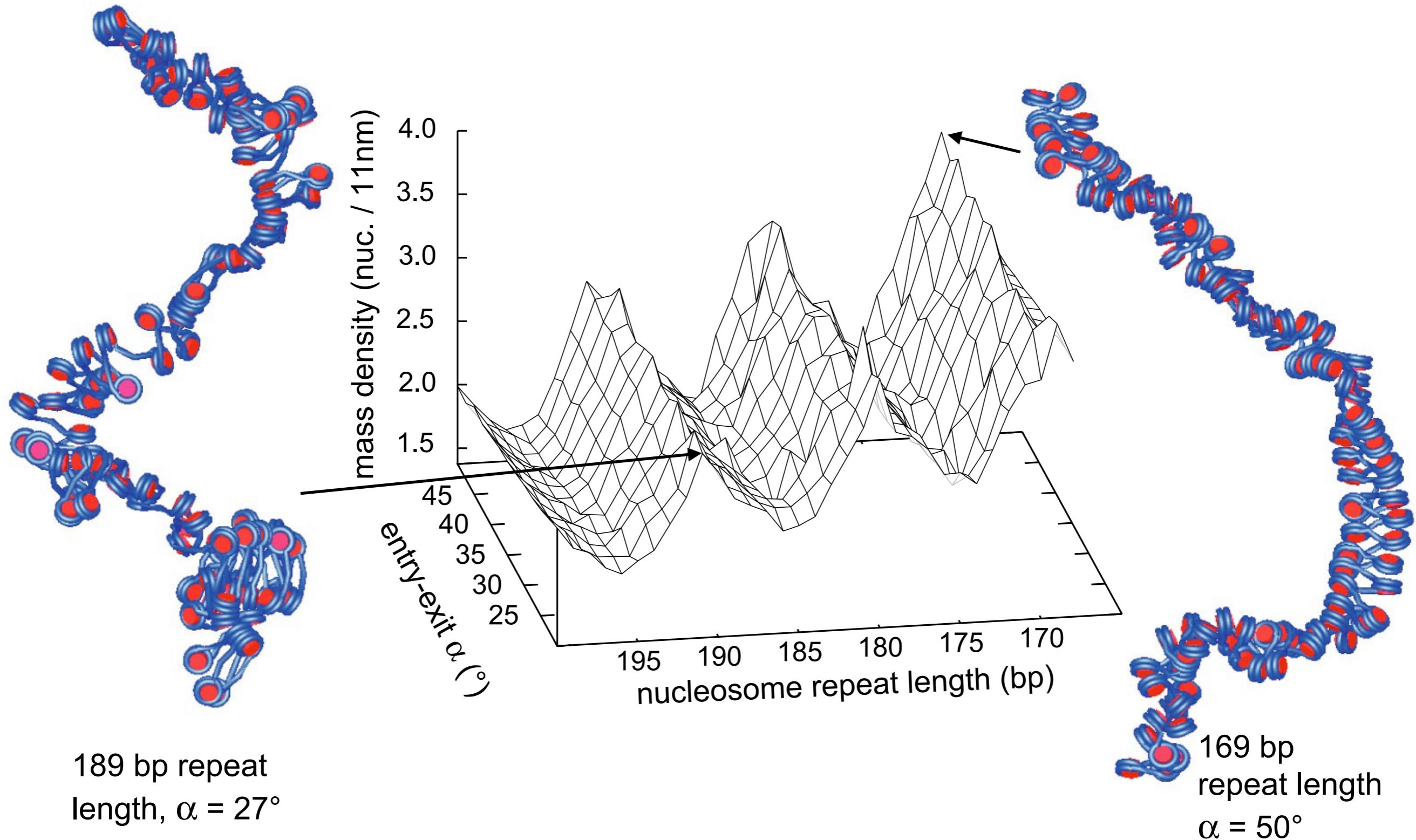
197bp x 61



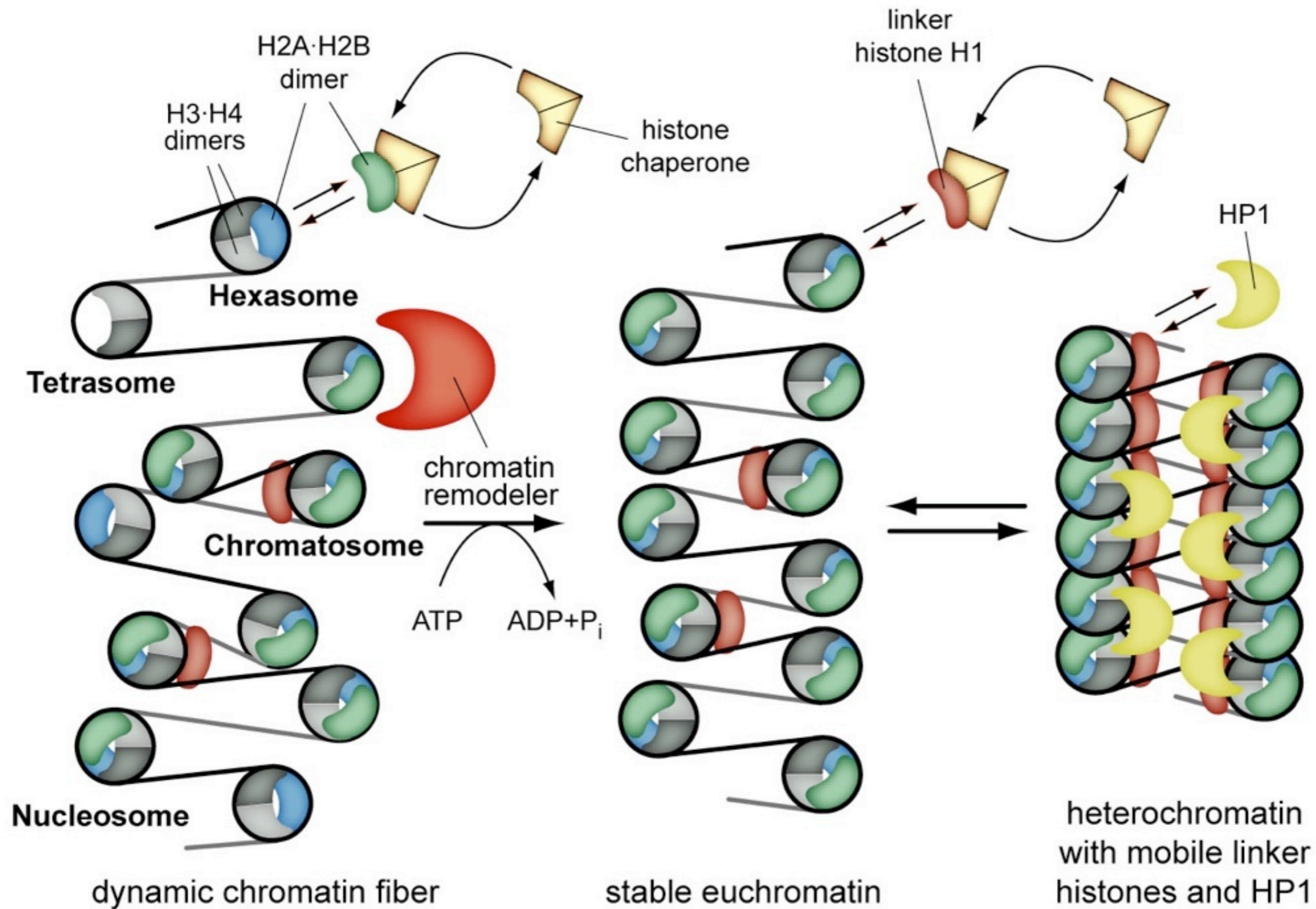
11.1 nuc/
11 nm fiber



CL fibers form at different repeat length with 10 bp periodicity and a mass density of 2.4-4 nucleosomes/11 nm fiber



Chromatin dynamics: histones and chromosomal proteins

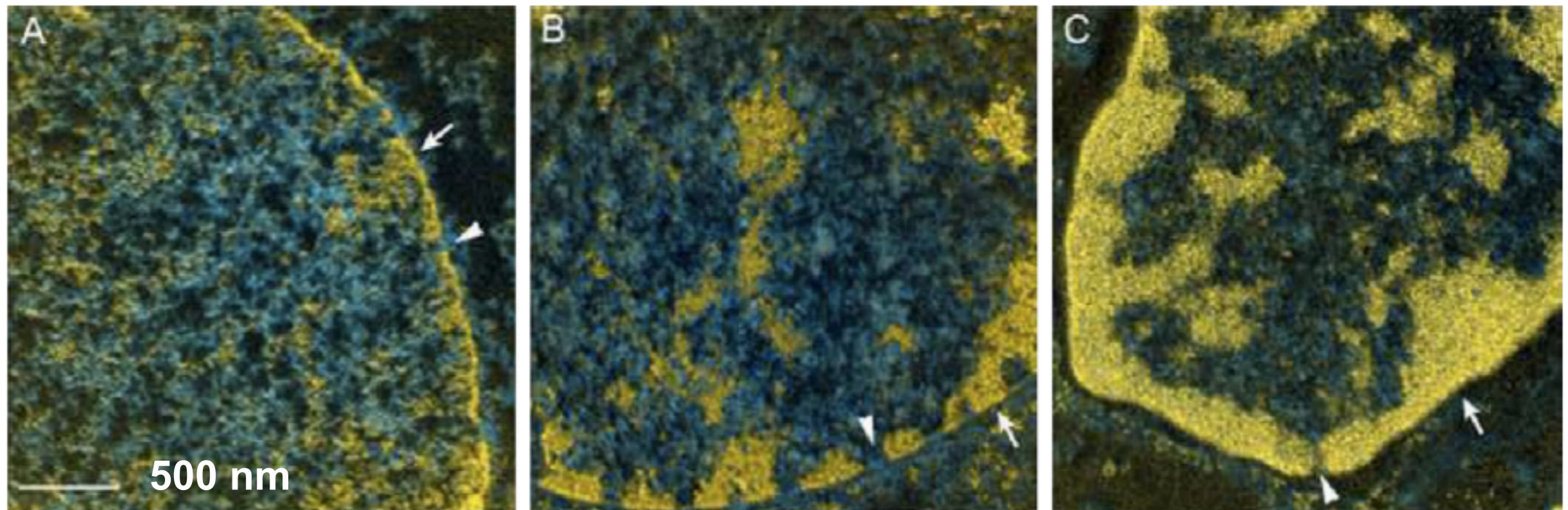


We know little about chromatin in the cell

Embryonic stem cell,
open chromatin

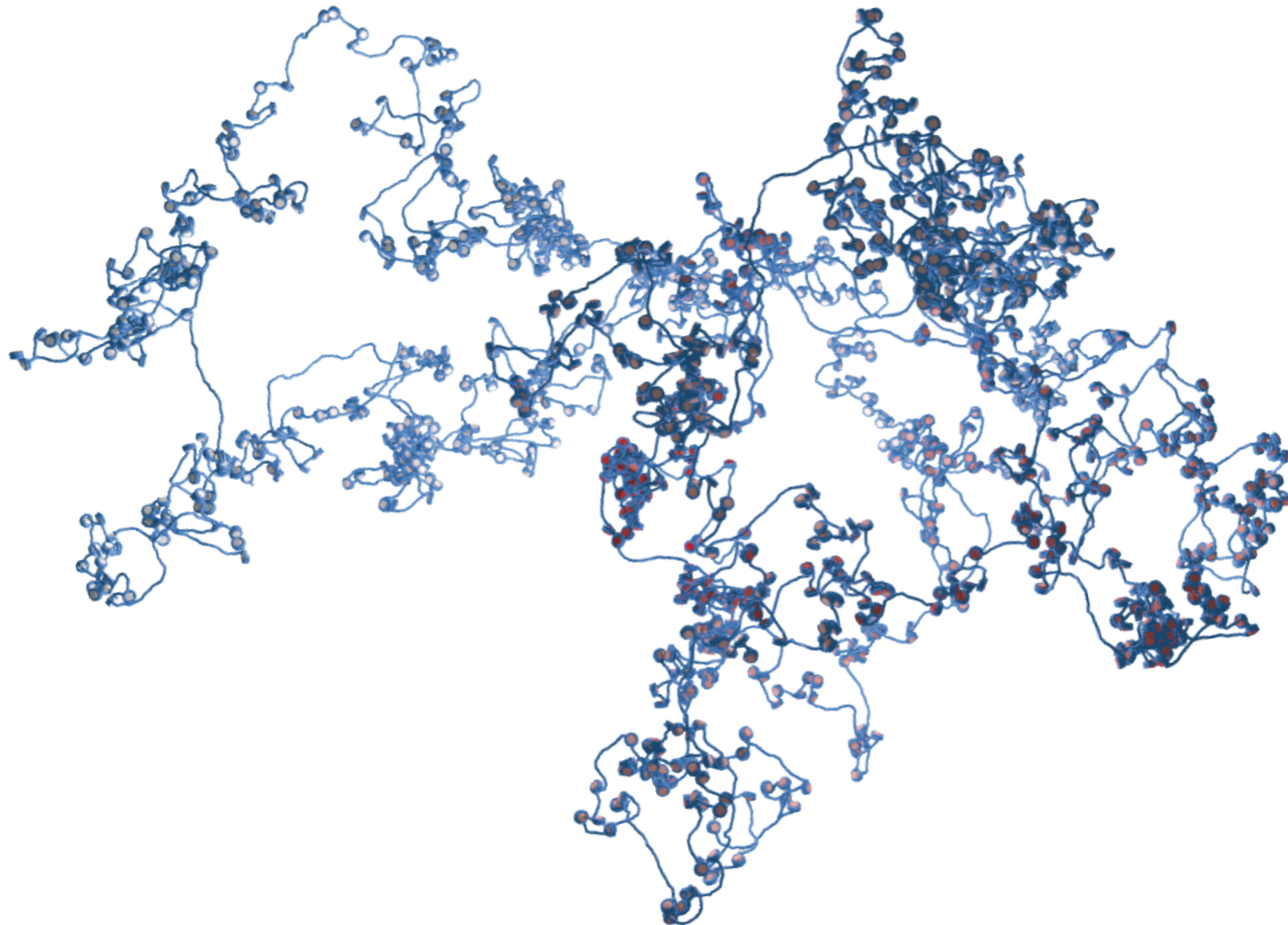
Hepatocyte cell,
intermediate compaction

Lymphocyte,
compact chromatin



ESI micrographs. In all images, chromatin is pseudo-colored yellow and protein-based structures cyan.

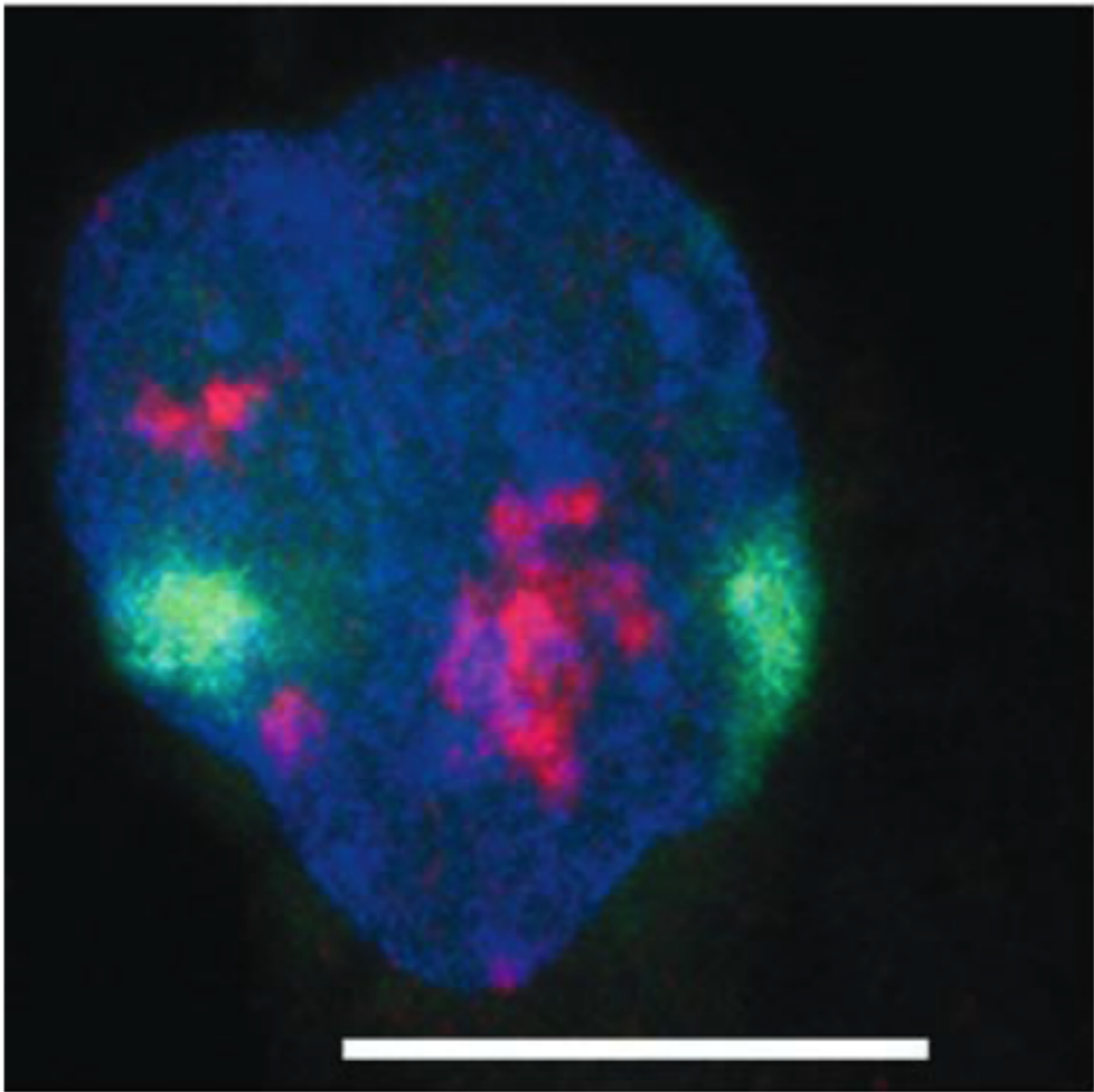
The nucleosome chain could be in a much more irregular conformation than concluded from in vitro experiments



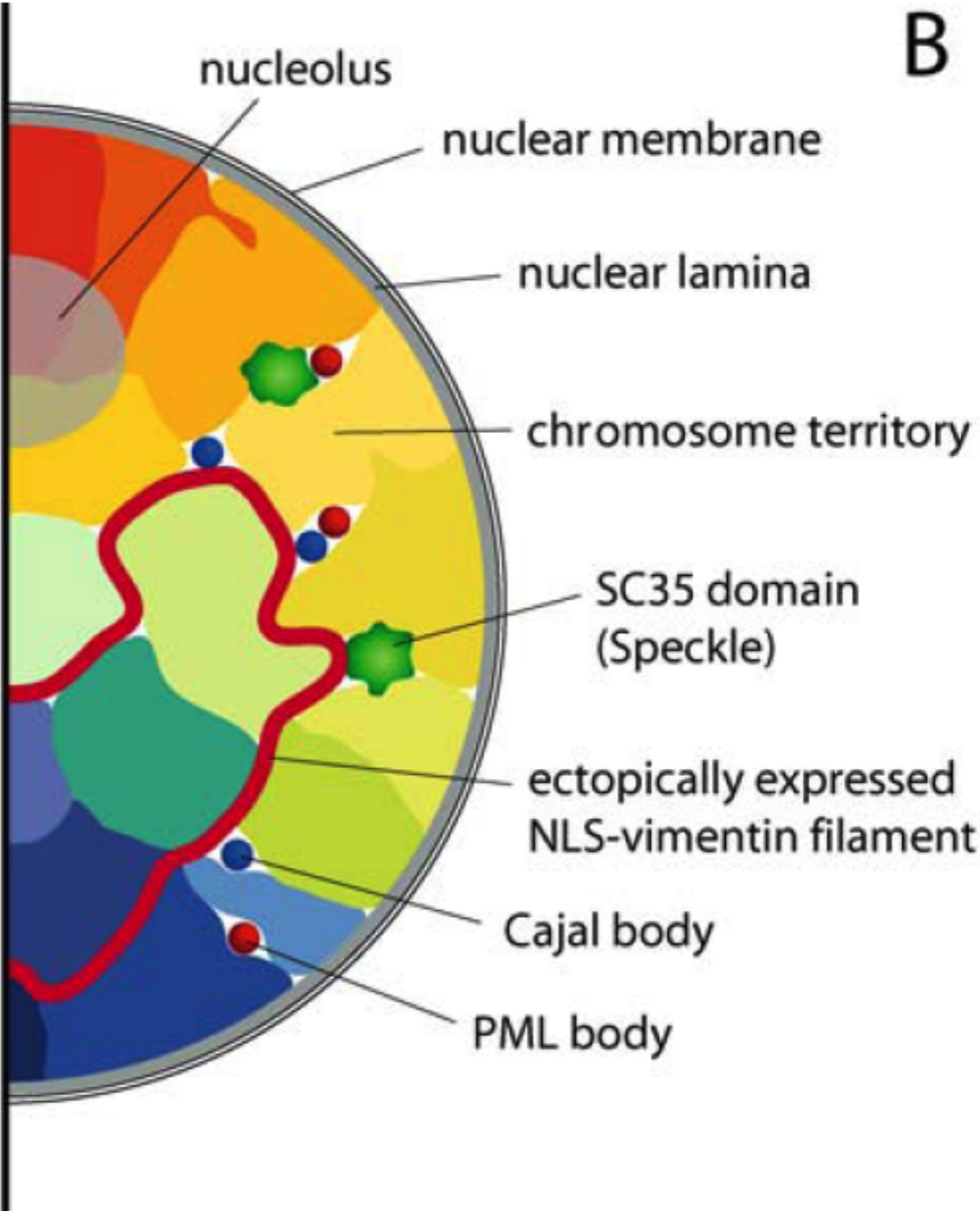
irregular spacing of nucleosomes, local heterogeneity, high concentration...

Chromosome territories

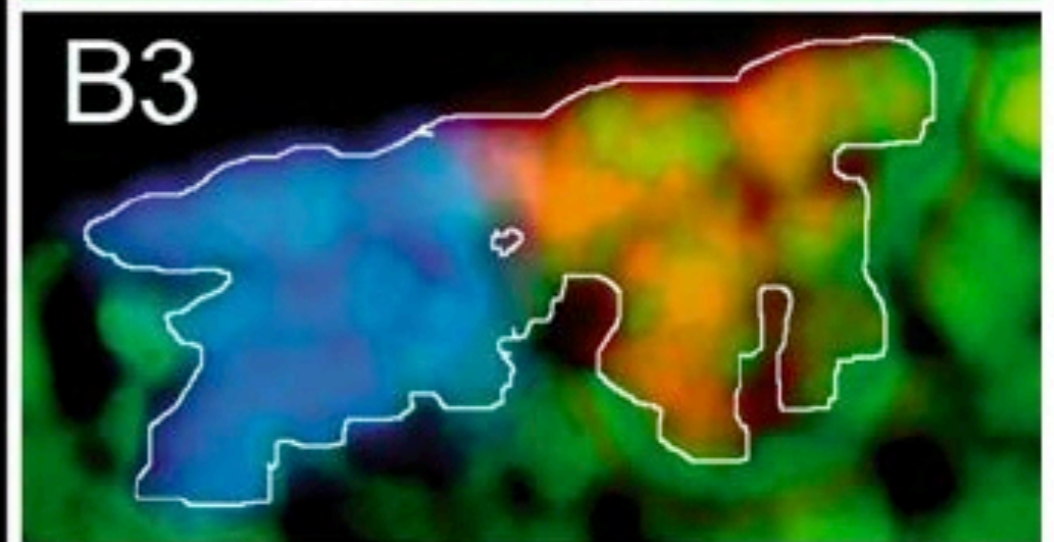
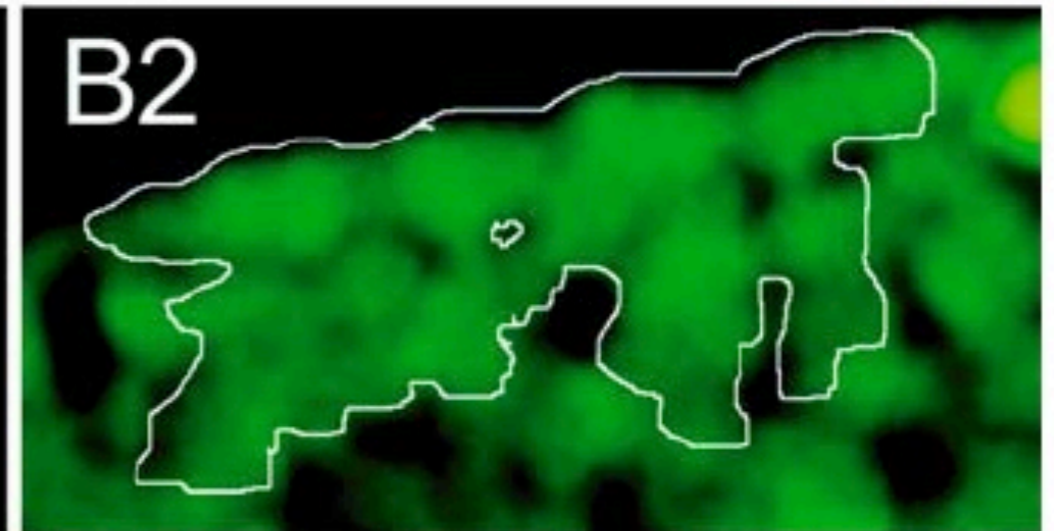
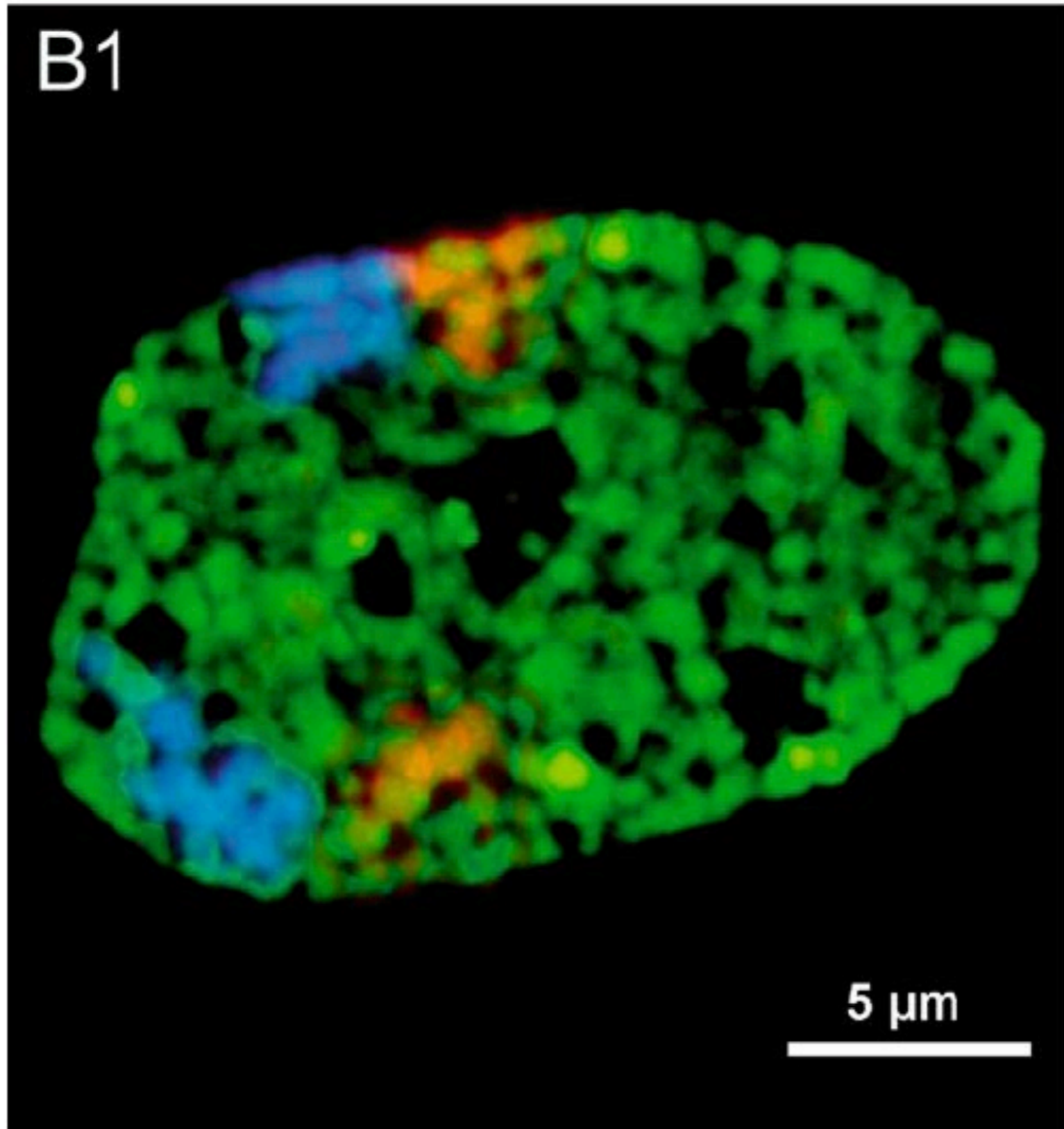
A



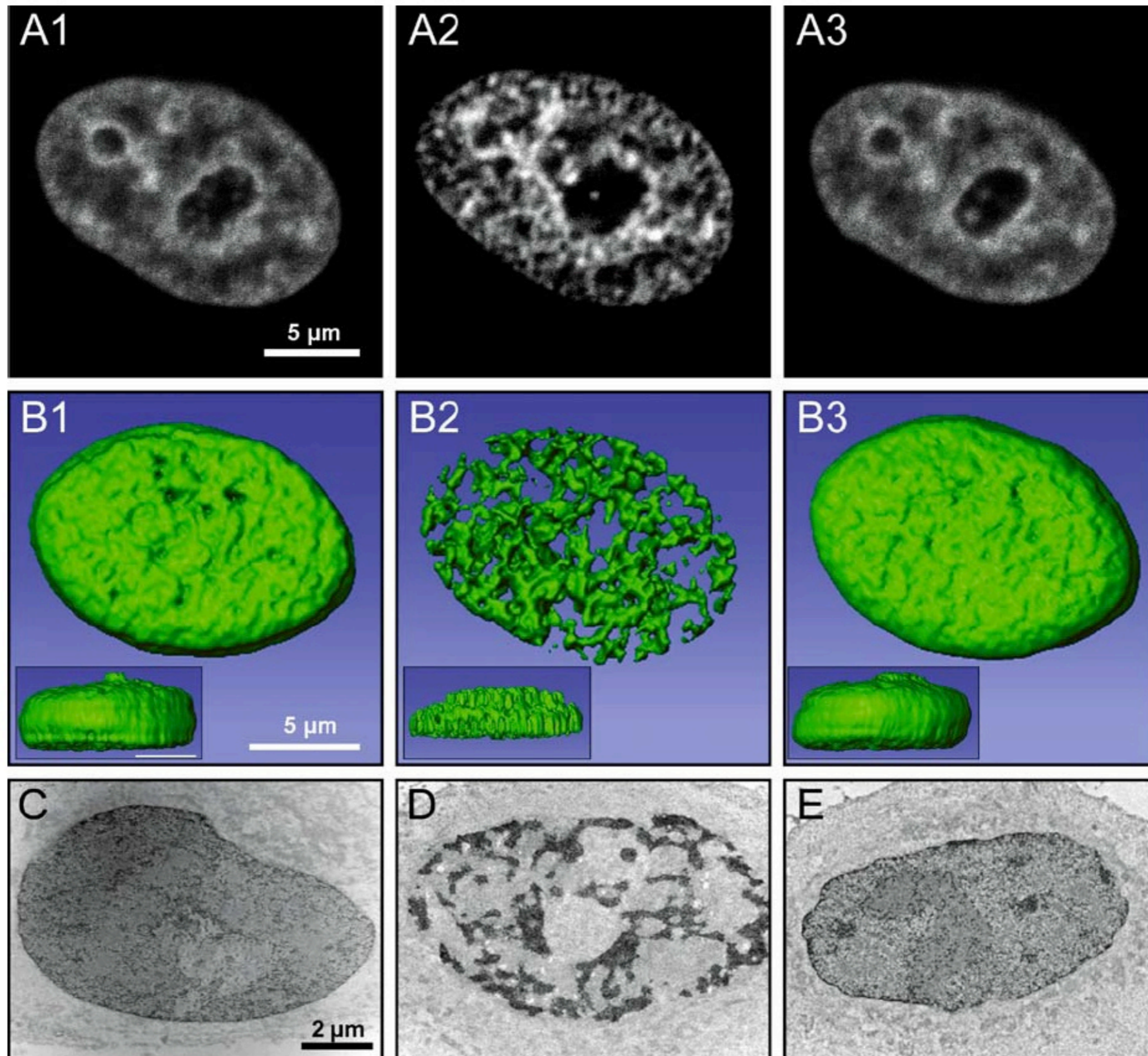
B



Chromosome territories (Albiez et al.)



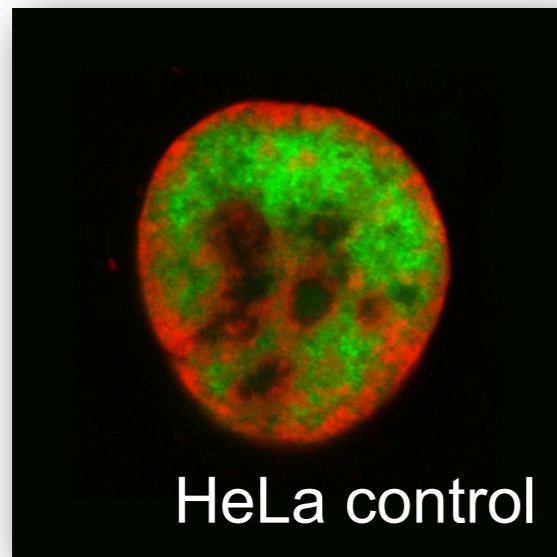
Chromatin can be reversibly condensed by increasing salt concentration in the medium or ATP depletion



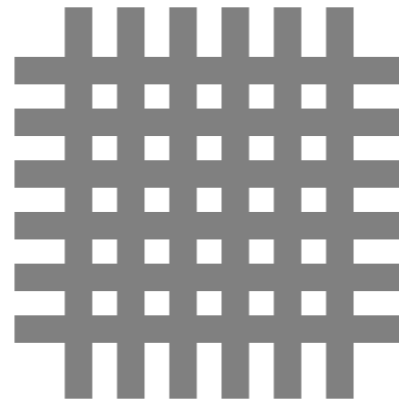
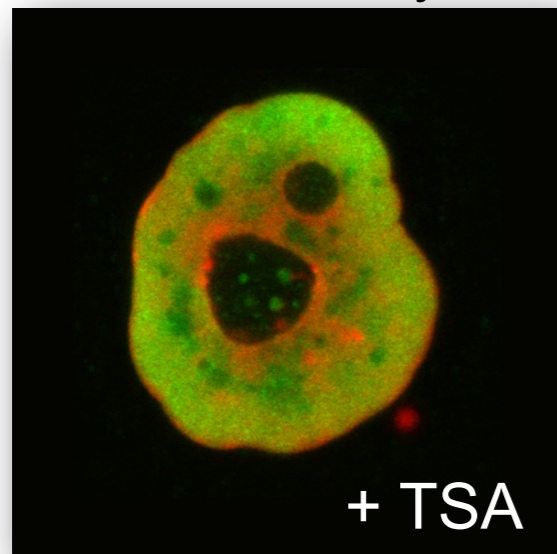
Injection of fluorescent dextrans (green) into the cell nucleus reveals chromatin (red) dependent accessibility

464 kDa dextran (green)
excluded from chromatin (red)

Results from analyzing the distribution of
42, 77, 148, 282, 464 and 2500 kDa dextrans

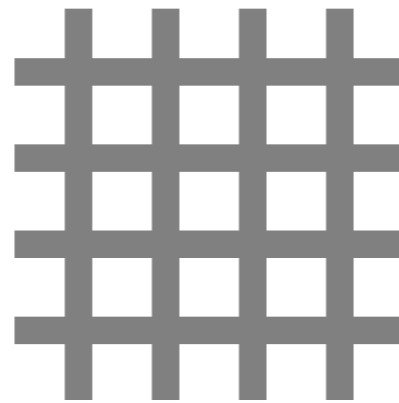


homogeneous
accessibility



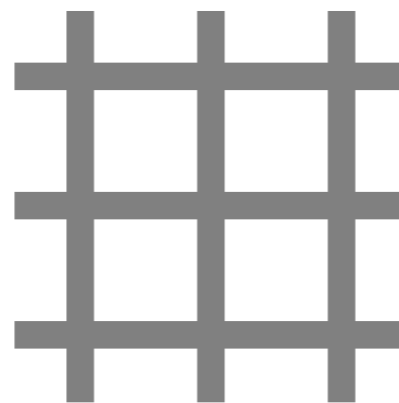
dense heterochromatin

- 16-20 nm pore size
- 0.4-0.5 mM nucleosomes
- accessible to proteins up to ~1 MDa



“bulk” heterochromatin

- 36-56 nm pore size
- 0.14-0.24 mM nucleosomes
- no restrictions to MDa mobile protein complexes but possibly to the formation active chromatin domains like 40-80 nm “transcription factories”



euchromatin or acetylated chromatin

- 60-100 nm pore size
- 0.06-0.13 mM nucleosomes
- excludes only large structure as PML/Cajal bodies (0.1-1 μm) into “interchromatin space”

Fejes Tóth et al. (2004) *J. Cell Sci.*, **117**, 4277-4287

Görisch, S., Lichter, P. and Rippe, K. (2005). *Histochem. Cell Biol.* **123**, 217-228.

Görisch, S., Wachsmuth, M., Fejes Tóth K., Lichter, P. and Rippe, K. (2005), *J. Cell Sci.* **118**, 5825-5834.

What's up next?

- Some basics on fluorescence to understand the techniques applied
- Looking at the inside of a cell from a protein's point of view
- Understanding how mobility and interactions of proteins are measured in living cells
- Dissecting epigenetic networks in living cells